Electron Microscopic Evaluation of Adhesion of Helicobacter pylori to the Gastric Epithelial Cells in Chronic Gastritis

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Background: The adhesion of H. pylori to the gastric epithelial cells may be an essential step for the pathophysiology of various H. pylori-induced gastrointestinal diseases. The purpose of this study was to investigate the ultrastructural relation of H. pylori and gastric epithelial cells in their adhesion.

Methods: Endoscopic biopsy of gastric antrum and body was performed from 15 patients (9 men, 6 women) with chronic gastritis and H. pylori infection. The specimens were processed for electron microscopy and observed with a transmission electron microscope (Hitachi H-600).

Results: On the basis of morphological appearances, the adhesions of H. pylori to the gastric epithelial cells were categorized into three types; filamentous connection, adhesion pedestals and membrane fusion. Coccoid and undetermined forms adhered mainly by the filamentous connection, whereas the bacillary forms adhered primarily by the adhesion pedestals and membrane fusion.

Conclusion: Various types of adhesion were associated with H. pylori and gastric epithelium. Further studies are needed to evaluate the influence of different types of adhesion to the pathophysiology of H. pylori.

Key Words: Helicobacter pylori: Gastric cell: Electron microscopy: Adhesion: Chronic gastritis

INTRODUCTION

Helicobacter pylori is a gram negative bacteria living in the stomach. It is typically curved bacilli in form, however the coccoid form is not rare^{1, 2)}. The bacillary form is known to be active in the pathogenesis of various gastric diseases, although the roles of coccoid form are not clear yet. As known so far, the bacillary form can convert to the coccoid form in bad environmental conditions such

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as lack of nutrients³⁾, increase of the intragastric pH⁴⁾, exposure to antibiotics⁵⁾, long-term culture⁶⁾, etc.

Ultrastructural studies have been used to elucidate the structural characteristics of *H. pylon*^{3, 7)} and to clarify the pathophysiologic association between *H. pylori* and infected gastric epithelial cells^{8, 9)}. Several investigators reported the adhesion of *H. pylori* to the gastric epithelial cells, suggesting such an adhesion may be closely related to the pathogenesis of *H. pylori*-related gastric diseases^{10, 11)}. Also, the frequent presence of coccoid forms near the severely damaged gastric epithelial cells suggests the morphological changes of *H. pylori* could be associated with the damages of the gastric epithelium^{10, 11)}.

This study aimed to evaluate the ultrastructural features of adhesion between the *H. pylori* and infected gastric epithelial cells as a way to understand the pathophysiology of *H. pylori*.

MATERIAL AND METHODS

1. Study Population

Fifteen patients (9 males and 6 females; average age 51) with chronic gastritis were selected for this study among those who visited the Department of Internal Medicine with upper gastrointestinal symptoms. They had no prior history of taking bismuth, H₂ antagonists, proton pump inhibitors or antibiotics for at least 4 weeks at their first visits to the hospital. The Urea Breath Test was performed and the presence of *H. pylori* was confirmed in the biopsy.

2. Electron Microscopy

Biopsies from the gastric antrum and corpus were performed during the diagnostic endoscopy of the upper gastrointestinal tract. The collected tissues were fixed in 2.5% glutaraldehyde, 0.1 M phosphate buffer (pH 7.4) and postfixed in 1% osmium tetroxide. After dehydrating with an ascending series of ethanol, the tissues were embedded in the EMbed-812. Sections were obtained with an ultramicrotome, double-stained with uranyl acetate and lead citrate and observed with H-600 transmission electron microscope (Nissei Sangyo, Japan) at the acceleration voltage of 75kV.

3. Analysis

All H. pylori attached to the surface of the gastric epithelial cells at a single thin section from each patient were photographed at $X10,000 \sim 12,000$. Firstly we analyzed the morphology of H. pylori and the types of adhesion between H. pylori and the gastric epithelial cells. Then, the adhesion was analyzed by counting the number of H. pylori according to the types of adhesion.

RESULTS

1. Morphology of H. pylori

Based on the appearance on the sections, *H. pylori* was categorized into three types: 1) a bacillary form with rod or curved shapes 2) a coccoid form with a round

Figure 1. A typical bacillary form of *H. pylori* (arrow) and several coccoid form (arrowhead) are noted. The cytoplasm is filled with heterochromatins with dark granules. Two undetermined forms of *H. pylori* are shown in lower insets. The cellular structure is not different from that of bacillary form.

shape and 3) an undetermined form with an intermediate morphology between bacillary and coccoid forms. No ultrastructural difference was observed between different types of *H. pylori* (Figure 1).

2. Adhesion Types

In this study, we defined the adhesion as an attachment of H. pylori to the gastric epithelial cells. Depending on the structures intervened, three types of adhesion were observed. In a filamentous connection, H. pylori was attached to the gastric epithelial cell via thin filaments (Figure 2). Adhesion pedestals; the elevated membrane thickening of gastric epithelial cells at the site of contact characterized some adhesion (Figure 3). Sometimes, H. pylori and the gastric epithelium were completely fused not to allow discretion, which was defined as a membrane fusion (Figure 4). Also, depending on the relative location of the H. pylori-attaching part with a reference to the virtual straight line connecting the superior border of the gastric epithelial cell, each type was divided into 2 patterns; 1) with membrane depression located below the line and 2) without membrane depression located above the line.

Figure 2. Examples of H. pylori attached to the gastric epithelium via filamentous connection (arrow). A bacillary form of H. pylori (A) is located on the surface and a coccoid form (B) is in the depression of the gastric epithelial surface.

Figure 3. A bacillary form (A) and a coccoid form (B) of *H. pylori* are attached to the gastric epithelial surface via adhesion pedestals. The attaching part of the epithelium is thickened and elevated (area between 2 arrows).

Figure 4. A bacillary form (A) and a coccoid form (B) of *H. pylori* are attached to the gastric epithelial surface directly. At the contact areas, membranes of *H. pylori* and gastric epthelium are fused (area between 2 arrows).

3. Relation of *H. pylori* and Adhesion Types
We summarized the observed number of different forms of *H. pylori* attached to the gastric epithelial cells depending on types of adhesion in Table 1. Coccoid and

undetermined forms were attached mostly by filamentous connections, whereas the bacillary form was attached mostly via adhesion pedestals and filamentous connections (Table 1).

Table 1. Number of H. pylori contacting gastric epithelium depending on the types of adhesion

	Filamentous connections		Adhesion pedestals		Membrane fusion	
	with depression	without depression	with depression	without depression	with depression	without depression
Bacillary form	3	14	0	20	1	5
Undetermined form	3	11	0	5	0	0
Coccoid form	4	18	0	10	3	1
Total	53		35		10	

DISCUSSION

Since the first identification and isolation of *H. pylori* from the human body by Drs. Marshall and Warren¹²⁾, *H. pylori* has been known to be a main factor for various upper gastrointestinal diseases, such as chronic gastritis, peptic ulcer, B cell lymphoma and gastric cancer. Many studies to elucidate the pathophysiologic roles of *H. pylori* are being progressed¹³⁾. Ultrastructural analysis with an electron microscopy can be a useful method of studies to observe the direct relationship of *H. pylori* to the gastric epithelium and to understand the pathologic changes of the gastric epithelium^{3, 8, 14, 15)}.

In an ultrastructural analysis, Smoot et al. 16) reported that in culture the degenerative changes of gastric epithelial cell membrane and the extinction of microvilli appeared with time. They also reported that H. pylori attached to the elevated part of gastric epithelial cells at the site of disappeared microvilli, which involves the actin polymerization¹⁶⁾. Similar involvement of actin polymerization in the adhesion was reported in E. coli's adhesion to intestinal epithelial cells¹⁷⁾. Different types of adhesion have been reported to be associated with the decrease in microvilli and the changes of cytoskeletons 1, 18, 19). It has been reported that glycocalyx is formed on the surfaces of gastric epithelial cells and H. pylori and mediates adhesion, especially in the form of filaments during the initial stage of adhesion, which is shown by lectin binding2, 201

The standard classification for adhesion types has not been suggested yet. Noach et al.80 classified the adhesion into 5 types; adhesive pedestals, membrane depression, abutting adhesion, membrane fusion and internalization, and reported the abutting adhesion as the most common type among them. During the analysis based on Noach et al's classification⁸⁾, we observed that the membrane depression and abutting adhesion were actually present, mixed with adhesion pedestals and membrane fusion. This led us to classify the adhesion from a new point of view. As written in the result section, we classified the adhesion broadly into 3 types, depending on the structures involved and subdivided each type into 2 patterns according to the membrane depression. The internalization in Noach et al's classification⁸⁾ was included in the pattern of "with membrane depression", because H. pylori observed in the gastric epithelial cells in this study were not present in the cytoplasm but in a vacuole-like structure thought to be a part of the deep invagination from the cell membrane. Putting aside the question whether *H. pylori* really invade the cell or not, the authors think that this pseudointernalization may be related to the same mechanisms that cause the membrane depression.

Even though direct relationship of different types of adhesion to the pathophysiology of *H. pylori* has not been proved, as far as we know, several evidences suggest the possibility that the different type of adhesion is not just a different morphological relation between *H. pylori* and the gastric epithelium, but a phenomenon showing a progress to cause some diseases^{1, 2, 11, 16, 18-20)}.

Hessey et al. 11) reported that the involution of cells was accelerated when the contact areas between H. pylori and gastric epithelium was wider. From this observation, they emphasized that the adhesion between H. pylori and the gastric epithelium should be understood as a pathophysiologic feature of H. pylori, not as a simple physical contact and suggested the possibility that H. pylori may accelerate the damage of the gastric epithelium through such types of adhesion as abutting and membrane depression¹¹⁾. Our results showed that H. pylori attaches the gastric epithelium most frequently by filamentous connections, followed by adhesion pedestals and membrane fusions. If Hessey et al's claim is correct, the adhesion pedestals without membrane depression in our report could be a beginning stage of the pathologic process leading to chronic gastritis. It is thought that further studies regarding the types of adhesion and the degree of cellular damage are necessary to make clear this point.

There are still arguments whether different morphology of *H. pylori* has any effect on the adhesion. Janas et al¹⁵⁾ reported the they observed no coccoid form at adhesion pedestals nor on any other adhesion sites. In this study, we observed lots of coccoid form of *H. pylori* adhered to the gastric epithelial cells, which was contrary to Janas et al's observation¹⁵⁾. It may be argued that many of the coccoid forms observed in this study can be artifacts due to sectional direction and we did not check this possibility by sectioning the specimen serially or examining by scanning electron microscopy. However, considering that almost the same numbers of attached coccoid and bacillary forms were observed in this study, we believe that a considerable number of coccoid forms attached to the gastric epithelial cells.

Our results showed that the coccoid forms adhere mainly by the filamentous connection, while the bacillary forms use adhesion pedestals and filamentous connection for adhesion. The adhesion of undetermined forms to the gastric epithelial cells was similar to the coccoid forms, in that they mainly attach to the gastric epithelial cells through the filamentous connection, but not through adhesion pedestals. Also, these undetermined forms have ultrastructural features not different from either bacillary or coccoid forms. Several reports suggested the ultrastructural changes of H. pylori during conversion from the bacillary to the coccoid forms 15, 21, 22). These suggest that the undetermined forms in our study contain a considerable number of intermediate forms, not only in shape but also in behavior, between bacillary and coccoid forms, as well as some obliquely sectioned bacillary forms. Even with the structural differences from earlier reports 15, 21, 22), the possibility that the undetermined form may represent a group of H. pylori changing their shapes from the bacillary into the coccoid still needs to be explored.

In conclusion, the results of this study suggest, based on the number of adhesion patterns observed, that the adhesion of *H. pylori* to the gastric epithelium might proceed in the order of the filamentous connection, adhesion pedestal formation and finally, membrane fusion. However, the possibility that different types of *H. pylori* may affect the gastric epithelium in different ways to form different adhesion structures, such as the filamentous connection by the coccoid form or the adhesion pedestal formation by the bacillary form, should be verified in further studies.

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REFERENCES

- Bode G, Malfertheiner P, Ditschuneit H. Pathogenic implications of ultrastructural findings in Campylobacter pylori related gastroduodenal diseases. Scand J Gastroenterol 23(Suppl 142):25–39, 1988
- 2. Jones DM, Curry A. Ther ultrastructure of Helicobacter pylori. In: Rathbone BJ, Heatley RV, eds. Helicobacter

- pylori and gastroduodenal disease, 2nd ed, Oxford, Blacwell Scientific Publication, p29-41, 1992
- Chan WY, Hui PK, Leung KM, Chow J, Kwok F, Ng CS. Coccoid forms of Helicobacter pylori in the human stomach. Am J Clin Pathol 102:503-507, 1994
- Moran AP, Upton ME. A comparative study of the rod and coccoid form of Campylobacter jejuni ATCC 29428. J Appl Bacteriol 60:103-110, 1986
- Berry V, Jenning K, Woodnutt G. Bactericidal and morphological effects of amoxacillin on Helicobacter pyton. Antimicrob Agents Chemother 39:1859–1891, 1995
- Moran AP, Upton ME. Factors affecting production of coccoid forms by Campylobacter jejuni on solid media during incubation. J Appl Bacteriol 62:527–537,1987
- Roe IH, Son SH, Oh HT, Choi J, Shin JH, Lee JH, Hah YC. Changes in the evolution of the antigenic profiles and morphology during coccoid conversion of Helicobacter pylori. Korea J Intern Med 14(1):9–15, 1999
- Noach LA, Rolf TM, Tytgat GNJ. Electron microscopic study of association between Helicobacter pylori and gastric and duodenal mucosa. J Clin Pathol 47:699–704, 1994
- Danielsson D. Ultrastructural study of two patterns in the interaction of Helicobacter pylori with neutrophils. J Clin Pathol 47:667–669, 1994
- Caselli M, Figura N, Trevisani L, Pazzi P, Guglielmetti P, Bovolenta R, Stabellini G. Patterns of physical modes of contact between Campylobacter pylori and gastric epithelium: Implication about the bacterial pathogenecity. Am J Gastroentero 84:511–513, 1989
- Hessey SJ, Spencer J, Wyatt JI, Sobala G, Rathbone BT, Axon AT, Dixon MF. Bacterial adhesion and disease activity in Helicobacter associated chronic gastritis. Gut 31(2):134–138, 1990
- Marshall BJ, Warren JR. Unidentified curved bacilli in the stomach of patients with gastritis and peptic ulceration. Lancet 16:1(8390):1311-1315, 1984
- Haas R, Burns BP, Asahi M: Pathogenesis of Helicobacter pylori. Current Opinion Gastroenterol 17 (Suppl. 1):S1-S5, 2001
- Caselli M, Aleotti A, Boldrini P, Ruina M, Alvisi V: Ultrastructural patterns of Helicobacter pylori. Gut 34: 1507–1509, 1993
- Janas B, Czkwianianc E, Bak-Romaniszyn L, Bartel H, Tosik D, Planeta-Malecka I: Electron microscopic study of Helicobacter pylori and gastric epithelial cells. Am J Gastroenterol 90(10):1829–1833, 1995
- Smoot DT, Resau JM, Naab T, Besbordes BC, Gilliam T, Bull-Henry K, Curry SB, Nidiry J, Sewchand J, Mills-Robertson K, Frontin K, Abebe E, Dillon M, Chippendale GR, Phelps PC, Scott VF, Mobley HLT. Adhesion of Helicobacter pylori to cultured human gastric epithelial cells. Infect Immun 16(1):350-355, 1993

H.J. Chun, D.K. Park, C.H. Park, J.H. Park, Y.T. Jeen, S.H. Um, S.W. Lee, J.H. Choi, C.D. Kim, H.S. Ryu, J.H. Hyun, Y.S. Chae, C.S. Uhm

- Knutton S, Baldwin T, Williams PH, McNeish AS. Actin accumulation at sites of bacterial adhesion to tissue culture cells: basis of a new diagnostic test for enterophathgenic and enterohemorrhagic Escherichia coli. Infect Immu 57(4):1290-1298, 1989
- Kazi JL, Sinniah R, Zaman V, Ng ML, Jafarey NA, Alam SH, Zuberi SJ, Kazi AM. Ultrastructural study of Helicobacter pylori associated gastritis. J Pathol 161: 65-70, 1990
- Thomson LL, Gavin JB, Tasman-Jones C. Relation of Helicobacter pylori to the human gastric mucosa in chronic gastritis of the antrum. Gut 13:1230-1236, 1990
- Rathbone BJ, Heatley RV. Helicobacter pylori and bastroduodenal disease, 2nd ed, Oxford, Blackwell Scientific Publications, p42-50, 1992
- Benaissa M, Babin P, Quellard N, Pezennec L, Cenatiempo Y, Fauchere JL: Changes in Helicobacter pylori ultrastructure and antigens during conversion from the bacillary to the coccoid form. Infect Immun 64(6):2331–2335, 1996
- Kusters JG, Gerrits MM, Van Strijp JAG, Vandenbroucke
 -Grauls CMJE: Coccoid forms of Helicobacter pylori are the morphologic manifestation of cell death. Infect Immun 65(9):3672–3679, 1997