

Macroaspartasemia as a Cause of Isolated Elevation of Aspartate Aminotransferase

- Its Biochemical and Physiological Characteristics -

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Objectives : *The increase of serum aspartate aminotransferase (AST) is generally found in hepatic, cardiac, muscular disease and hemolytic disorders of the red blood cell (RBC). The elevation of its activity is suspected in pathological conditions of these organs. However, instances without any of those conditions rarely exist.*

Methods : *The experimental samples were obtained from a normal person's hemolysed RBC, a hepatitis patient and a macroaspartatemic female's serum. They were studied with exclusion chromatography, electrophoresis of AST and changes of AST activity due to Polyethylene Glycol (PEG) and various conditions on storage.*

Results :

1) *The patterns of AST activity by exclusion chromatography are similar to the hemolysed RBC and the hepatitis's serum but differs by the isolated AST elevation.*

2) *The AST activity with addition of PEG and different anti-immunoglobulin subtypes to different serums are slightly decreased in hepatitis but markedly decreased with PEG and anti-IgG in macroaspartatemia.*

3) *The patterns of AST activity in electrophoresis are single band - cytosomal AST (cAST)-from hemolysed RBC and two bands-mitochondrial AST (mAST) and cAST-from hepatitis, the major being cAST and the minor mAST. Even though there are two bands, the major one is atypical and the minor corresponds to mAST in macroaspartatemia.*

4) *The changes of AST activity on storage according to time and temperature show to be stable over 4weeks at room temperature and cooled condition, and 9weeks under frozen state in macroaspartatase.*

Conclusion : *Concluding from the above findings, macroaspartatemia is an enzyme-immunoglobulin complex composed of cAST with IgG. MacroAST might be stabler than usual AST at physical conditions.*

Key Words : *Macroenzyme, Isoenzyme, Aspartate aminotransferase*

INTRODUCTION

The enzyme of aspartate aminotransferase (AST) is present in a wide variety of tissues-including heart, skeletal muscle, kidney, red blood cell (RBC)

and brain, in addition to liver¹⁾. So the elevation of AST is suspected of being due to injury of the above mentioned organs. Even in the injury of these, they usually are associated with other abnormalities of enzymes or metabolites such as elevation of creatine kinase (CK) and lactic dehydrogenase (LDH) in myocardial infarction. An isolated and persistent elevation of AST occasionally can be found in advanced hepatocellular carcinoma, alcoholic liver disease and to some drug

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effects in hepatic disorders²⁻⁴, but these instances are very rare without the above mentioned conditions⁵⁻¹⁰. We experienced one case of the above condition and determined to study it.

CASE REPORT

A 24-year-old woman visited our department for evaluation of hepatic function because of isolated AST elevation. It had begun two years before. She had no association with any other symptoms. She denied any alcohol use, smoking and drug medication. Her family histories were non-specific except that her father had labile hypertension. Findings on physical examination were unremarkable.

The results of all laboratory studies were normal, except for an unexplainable elevation of AST at 196IU (normal 16-40). The results of radiological examinations, such as plain chest film, liver scanning, ultrasonographic and computer tomographic finding of abdomen, were unremarkable. She was considered as an unusual "hyperaspartemia syndrome" and recommended for interval check of hepatic functions.

5 months later, she revisited our department for accurate evaluation of hepatic function. The results of all laboratory studies were normal, except for isolated AST elevation at 223IU (normal 16-40). Her physical status was as follows: height 160cm, 53Kg, 100/60mmHg. Results of other liver function tests, including bilirubin, albumin, prothrombin time, alkaline phosphatase (ALP) and ALT were normal. On repeated testing, laboratory results, AST was 217IU, ALT 13IU, ALP 39IU, LDH 71IU (N:53-137), creatinine 0.9mg/dl, blood urea nitrogen (BUN) 11mg/dl, CK 59IU (N:60-103) and total bilirubin 0.6mg/dl. All serological marker for hepatitis B, C and E virus were negative. Anti-body of IgG to hepatitis A virus was positive, but IgM was negative. The total protein, serum iron, transferrin, CBC, electrolyte, glucose and thyroid functions were all normal. Serological examination for rheumatoid arthritis (RA) factor, LE cell, ANA, AMA and anti-smooth muscle antibody were negative. Special studies confirmed that the patient had an immunoglobulin-complexed AST.

MATERIALS AND METHODS

1. Subjects and material

The samples of our study were collected from a patient with acute viral hepatitis (AVH: due to HBV), the above mentioned female and purified cytosolic enzyme from hemolysed RBC. The cytosolic AST was purified from a normal person's RBC as previously described by Rej et al¹¹.

2. AST Assay

Routine and our one case of elevated AST determinations were performed as the chemistry part of the clinical pathology on a Spectrum EPX (Abbott[®], IL).

3. Exclusion Chromatography

Exclusion chromatography was done with each 1-ml serum samples, including our isolated AST elevation case, AVH patient and the above mentioned purified cAST. The column used was 1.5-by 45-cm column of Sephacryl G-300 (Pharmacia, NJ). The elution was performed with 0.01M Tris buffer and 0.15M NaCl buffer (pH 7.2) at a flow rate of 15ml/h. Column fractions were assayed directly for AST assay as described in the above paragraph.

4. Electrophoresis

Universal agarose films and buffer were purchased from Ciba-Corning Medical (LA). The samples (1.2ml) of AVH patient's and subject's serum and purified cAST were applied to each well. These samples were electrophoresed for 35 minutes at 100volts in universal barbiturate buffer (pH 8.6).

5. Immunoglobulin electrophoresis

When the electrophoresis was complete, the plates were laid and filled with anti-human immunoglobulin complex, G, A, M, GMA, kappa and lambda between the gaps. Then these were incubated 24 hours at room temperature and stained.

6. AST electrophoresis

AST staining was done as previously described by Sakakibara et al¹². To briefly describe it, after the electrophoresis was complete, the plates were layed with a film of 1ml of the AST reagent mixture [10umol alpha-ketoglutarate, 200umol L-cysteine sulfinate, 0.1mg m-PMS, 0.8mg MTT, 2umol EDTA, 20mg dextran and 100umol imidazole buffer (pH 7.5)] for 20minutes of incubation at 37°C. Then the agarose plates were immersed in 10% acetic acid solution for 5minutes, washed with water and then dried in an oven at 65°C. AST activity was visualized by the appearance of purple bands.

7. Polyethylene Glycol (PEG) precipitation and Identification of Antibody class

As the methods of screening examination of enzyme-immunoglobulin complex and identification of its antibody subclass, we used the methods with slight modification previously described by Litin et al⁵. Briefly described in our method, PEG 6000 was purchased from Sigma chemical company. We added 100ul of a 24% solution of PEG 6000 to 100ul of serum from each subject. A blank was prepared by substituting 100ul of phosphate buffer solution (PBS) for PEG. The mixtures were incubated at 37°C for 10 minutes and centrifuged at 3,000×g at room temperature for 20 minutes. AST activity was assayed from their supernatants, as previously described. The identification of the immunoglobulin associated with AST was done by selected precipitation with rabbit anti-human antiserum. To 20ul of subject serum was added 180ul of the antiserum and then shaken the mixture was shaken gently and incubated at room air for 60 minutes. After incubation, the mixture was centrifuged at 3,000×g for 15minutes. AST activity was assayed from their supernatants as previously described.

8. Changes of AST activity according to temperature and time on storage

Each serum sample was divided into 3parts according to interval and stored at frozen state (-20°C), cooling state (4°C) and room temperature. Then the AST activity was assayed as previously

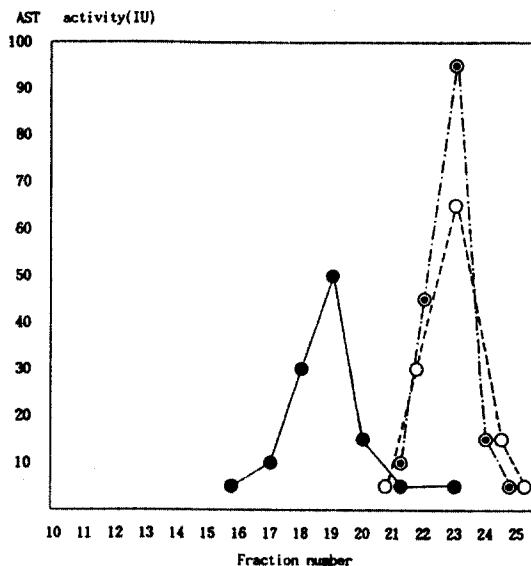


Fig. 1. Gel exclusion chromatography (on Sephacryl G-300) of serum specimens from an acute viral hepatitis patient (○) 25 year-old-male, due to hepatitis B virus and isolated AST elevation female (●) described in this report and hemolysed RBC (◐). AST activity was determined in each of the fractions.

described.

RESULTS

1. Exclusion Chromatography

The patterns of AST activity were apparently different from the isolated hyperaspartemic subject and purified cAST and AVH patient. The patterns of AST activity were similar with purified enzyme from hemolysed RBC and AVH patient, but different with isolated hyperaspartemic subject. The AST activity was detected in the earlier fraction of the isolated hyperaspartemic subject and not in the other two groups (Fig. 1). This implies that it is a macromolecule with higher molecular weight.

2. Electrophoresis

1) **AST electrophoresis**: There was a single band from hemolysed RBC which shows in pure cytosomal type of AST (cAST) activity, but there were two bands in the AVH patient and the isolated AST elevation case. There were two

MACROASPARTASEMIA AS A CAUSE OF ISOLATED ELEVATION OF
ASPARTATE AMINOTRANSFERASE

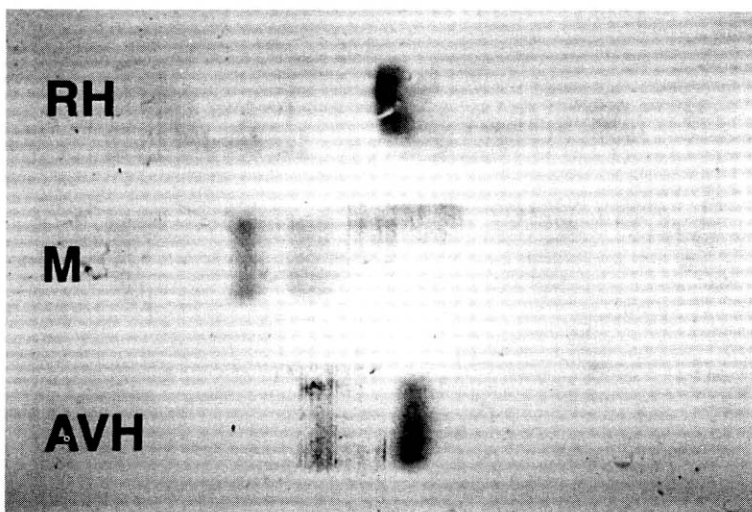


Fig. 2. The electrophoresis patterns of samples [lane RH: from purified aspartate aminotransferase (c-AST) from hemolysed red blood cell (RBC), lane AVH: serum from acute viral hepatitis patient due to hepatitis B *vivire*, lane M: serum from macroaspartatemia patient] after AST stain by Sakakibara's method. There is single band in RH lane (purified cytoplasmic-AST from RBC: 250IU). In lane AVH, there are two bands of which one is a major band corresponding to c-AST in RH lane and the other is a minor band suggested to mitochondrial-AST (m-AST). In M lane, there are two bands of which one is a minor band suggested to m-AST in AVH lane and the other is a major band which is a new one. See text for further discussion.

bands in the latter group, but the pattern differed with each. In the AVH patient's serum, there were 2 bands of which the thick band corresponded to cAST activity and the thin one to mitochondrial (mAST) type. This finding means that the major portion of circulating AST activity in AVH patient is a cytosomal type. In the macroaspartatemic case, we can find a thin band corresponding to mAST without cAST activity, but instead of the cAST type we can find abnormal thick AST activity which is near to the slit of the start, which is similar to the characteristic pattern of those previously mentioned macroaspartatemia (Fig. 2). This finding suggests that this case of hyperaspartatemia is due to mitochondrial type and abnormal type of AST activity. The abnormal type of AST activity might be cytosomal type of AST due to other effects.

2) Immunoglobulin electrophoresis: The no abnormal pattern in the AVH patient (not

shown), and there was only one abnormality in immunoglobulin G (Fig. 3).

3. Polyethylene Glycol (PEG) precipitation and Identification of Antibody class

PEG is well known to precipitate antibody-antigen complexes and has been suggested as a rapid screening test for macroaspartatemia. Because of this, precipitation experiments with PEG were undertaken in an AVH patient and a macroaspartatemic patient. The rates of AST activity from each supernatant were different. The AST activity was markedly dropped from 223IU to 23IU in the macroaspartatemic patient, but slightly dropped from 430IU to 378IU in the AVH patient. It is also well known as a rapid screening test for the identification of antibody class by addition with the anti-immunoglobulin class in macroaspartatemia. Identification of antibody class was done by selected precipitation with rabbit anti-human anti-

anti-TIg
 anti-IgG
 anti-IgA
 anti-IgM
 λ
 K
 anti-IgGAM

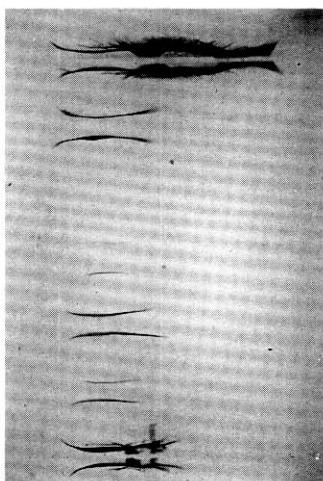


Fig. 3. Immunoglobulin electrophoresis patterns of macroaspartemic patients. There is an atypical precipitin arc in the total anti-immunoglobulin (anti-TIg), anti-immunoglobulin G (anti-IgG) and anti-immunoglobulin GMA complex (anti-IgGMA). But there is not an atypical precipitin arc in the anti-immunoglobulin A and M. See text for further discussion.

Table 1. The Changes of AST (IU) Activity from the Supernatants with Precipitation by Addition of Polyethylene Glycol (PEG) and Rabbit Anti-human to IgG, A and M

	macro-AST case	AVH patient
original serum	223	430
H ₂ O(blank)	230	434
PEG	23	378
anti-IgG	28	388
anti-IgA	198	396
anti-IgM	188	346

serum Ig G, A and M. There were slight changes of the AST activity in the AVH patient, but marked changes in the macroaspartemic patient, especially to immunoglobulin G from 223IU to 24IU (Table 1). Even if there was any change in its activity, it was not significant as anti-immunoglobulin G. So the change of AST activity might be specific to Ig G in our macroaspartemic patient.

Table 2. Changes of AST (IU) Activity According to Time and Temperature by Various Condition of its Storage

day	0	3	7	10	14	21	28	42	72
AVH RT		423	412	410	418	342	NS	NS	NS
AVH 4°C	430	434	428	416	412	NS	NS	NS	NS
AVH -20°C		424	430	416	418	NS	NS	NS	NS
MAST RT		217	220	212	208	207	210	208	167
MAST 4°C	223	228	224	214	220	212	216	213	181
MAST -20°C		218	227	210	207	216	215	208	212
RH RT	562	546	538	552	NS	NS	NS	NS	NS

Legends are as follow: AVH; acute viral hepatitis, MAST; macroaspartemic patient, RH; hemolysed red blood cell (RBC), RT: room temperature

4. Changes of AST activity according to temperature and time

To evaluate the characteristics of macro-AST due to any variable condition, AST activities were assayed in variable conditions. We could not assay enough because of deficiency of serum. There were no changes of AST activity due to purified enzyme from hemolysed RBC for 10 days, hepatitis serum for 14 days, in macroaspartemic patient for 6 weeks in room temperature and cooled condition, and 9 weeks in frozen state (Table 2).

DISCUSSION

AST is a well-known enzyme found in a wide variety of tissues—including heart, skeletal muscle, kidney, RBC and brain, in addition to liver. So this elevation is suspected of being due to the injury of the above mentioned organs. The injury is usually associated with other abnormalities of enzymes or metabolites such as elevation of bilirubin, alkaline phosphatase (ALP), alanine transaminase (ALT) and lactic dehydrogenase (LDH) in hepatic injuries due to their etiologies⁹⁻¹⁰. Even if there is an AST elevation without ALT elevation in advanced hepatocellular carcinoma, alcoholic liver diseases and hepatic disorder with drug effect, they usually are associated with other abnormalities of enzymes or metabolites and their abnormality can be found with other diagnostic stools such as history and radiological methods. However,

MACROASPARTASEMIA AS A CAUSE OF ISOLATED ELEVATION OF ASPARTATE AMINOTRANSFERASE

there rarely exist instances without those pathological conditions.

The usual method for approximating the size of an enzyme while retaining its activity is by exclusion chromatography. In this technique, larger proteins are excluded from the pores of the resin and, therefore, appear in the earlier fractions than do smaller proteins. It is even possible to approximate the molecular size by exclusion chromatography. In this reported case, isolated AST elevation caused clinicians to suspect liver disease, despite unrelated finding in the history and on physical and laboratory examinations. Liver and skeletal muscle biopsy specimens were not obtained to rule out the disease state in our case, microscopically, and were extremely unlikely because of normal CK, LDH, ALT and creatinine. Our chromatographic results easily realized those AST activities of purified enzyme from RBC, and hepatitis patients are an identical pattern, but differ to the isolated AST elevation case. It also appears in the earlier fraction in an isolated AST elevation case. By addition to these findings and rapid screening test by PEG precipitation, our case can be diagnosed as macroaspartatemia composed with enzyme-immunoglobulin complex⁸.

There are two circulating AST isoenzymes in the serum—one is cytoplasmic and the other is mitochondrial¹³ in their origin. The isoenzymes can be easily separated by electrophoretic or immunochemical methods. Even the mitochondrial isoenzyme is a major part during some severe tissue injury¹⁴, usually circulating serum AST activity which arises from the cytoplasmic isoenzyme. In most of the reports about macro-AST, they mentioned it is a enzyme-immunoglobulin complex especially due to Ig G or rarely Ig G associated with IgA, but did not mention which kinds of AST isoenzyme type were combined with immunobulin, except one^{5, 8, 10, 15}. PEG is well known to precipitate antibody-antigen complexes and has been suggested as a rapid screening test for macroaspartatemia^{5, 9}. So, our case might be an AST-immunoglobulin complex. In our case, the macro-AST complex might be an immunoglobulin-cytoplasmic AST isoenzyme, as the generally circulating AST isoenzyme is cAST¹³⁻⁴. Also in our AST electrophoresis, AST activity was not obser-

ved as cytoplasmic type but observed only a abnormal type of AST and mitochondrial type. In one report about immune-complexed AST obtained biopsy material from the liver and skeletal muscle⁹, the electrophotogram was similar to our study. AST isoenzymes from these tissues were normal. As circulating serum AST activity is a major fraction of the cytoplasmic type, it is suggested that normal cytoplasmic AST isoenzyme is bounded to immunoglobulin only after entry to the circulation^{9, 14-5}. This evidence was confirmed later by Stasia et al with monoclonal antibody¹⁰.

The type of immunoglobulin associated with macro-AST complex can be easily identified by selected precipitation with PEG and anti-serum Ig G, A and M as a rapid screening test, as mentioned previously by Litin et al⁵. But with this method, AST activity was mildly reduced due to non-specific binding of immunoglobulin. In our case of the macro-AST, the AST activity was markedly reduced especially to immunoglobulin G. Even though it was slightly changed in the hepatitis patient and more decreased to anti-immunoglobulin A and M, it was not significant as anti-immunoglobulin G. Also there was an abnormal finding of IgG in immunoglobulin electrophoresis. So, the change of AST activity might be specific to Ig G in our macroaspartatemic patient.

A recent review on serum AST pointed out the equivocal and conflicting reports on the effects of its storage¹⁶. Generally AST activity in serum or plasma was stable within 2 weeks of storage at 4°C, 4 weeks at -20°C and in vitro frozen state lyophilized purified AST activity may be maintained for 1 year with minimal loss of its activity¹⁷⁻⁸. In our case of macroaspartatemia, there was no change of AST activity for 4 weeks at room temperature and cooled condition, and 9 weeks in frozen state. This is the first report about the AST activity on the effect of its storage in various condition. Although we can not confirm that the enzyme activity of macro-AST might be maintained longer than normal AST, this evidence may suggest that the enzyme activity of macro-AST might be more stable than normal AST.

The causes of the increased enzyme activity in serum for a long time are not suggested. The mechanism for this increased activity is best

understood for macroamylase. Amylase (half life : about 2hr.), a low molecular weight protein (MW: -55,000Da), is readily filtered and excreted by the kidney, whereas macroamylase has high molecule and is not filtered by kidney¹⁹⁻²¹. But aspartate aminotransferase (EC 2.6.1.1-100,000Da) is larger than amylase. Also, the clearance of AST is not clearly defined, but its fractional catabolic rate constant from plasma is $0.088 \pm 0.016 \text{ hr}^{-1}$ (half life : about 23hr.)²². Usually the molecular mass of immune-enzyme complexed as determined by gel filtration and ultracentrifugation are consistent with a two-to-one ratio between enzyme and antibody. Also, the antibody (especially IgG: -180,000Da) has a larger molecule and is clear by the reticuloendothelial system (RES)^{21, 23-5}. It has been proposed that antibody binding to serum enzyme interferes with RES clearance mechanism. As mentioned in the above reasons, macro-AST complex might be cleared by RES with interference, so its activity might be higher in the serum.

Several enzymes complexed with immunoglobulin have been described and this group of immunoglobulin-complexed enzyme disorders has been termed the "ICE disorders"²⁶. These conditions seem to resent a nonspecific dysproteinemic response of the human immune system. Until now, the known immunocomplexed enzymes are amylase, CK, LDH, ALP, ALT, glucose-6-phosphate dehydrogenase, acid phosphatase, lipase and AST^{20, 23, 25, 27-31}. Several corresponding clinical conditions and their enzyme activity, related to their disease activity, have coexisted with these enzyme disorders, although any relationship between disease states and ICE disorders is purely speculative²⁴. In cases of macro-AST complex, it is known that the conditions associated with macro-AST complex are cardiovascular, gastrointestinal, endocrinologic, autoimmune and infectious diseases³⁰⁻¹. In our case, we can not find out any associated disease.

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REFERENCES

1. Stolz A, Kaplowitz N. *Biochemical tests for liver disease*. In Zakim D, Boyer TD *Hepatology: A textbook of liver disease*. 2nd ed p.652, Philadelphia WB. Saunders Co, 1990.
2. Shimokawa Y, Okuda K, Kubo Y, et al. *Serum glutamic oxaloacetic transaminase / glutamic pyruvic transaminase ratios in hepatocellular carcinoma*. *Cancer* 1977; 40:319-324.
3. Cohen JA, Kaplan MM. *The SGOT/SGPT ratio-an indicator of alcoholic liver disease*. *Dig Dis Sci* 1979; 24:835.
4. Lee HS, Kim YT, Jung HC, et al. *Prospective, randomized, controlled trial with diphenyl-dimethyl-dicarboxylate in chronic active liver diseases: The effect on lowering serum alanine aminotransferase levels*. *Korean Journal of Medicine* 1991; 41:172-178.
5. Kontinen A, Murros J, Okada K, Salaspuro M, Somer H, Ra'sa'nen J. *A new cause of increased serum aspartate aminotransferase activity*. *Clin Chim Acta* 1978; 84:145-147.
6. Weider N, Lott JA, Yale VD, Wahl RL, Little RA. *immunoglobulin-complexed aspartate aminotransferase*. *Clin Chem* 1983; 29:382-384.
7. Moriyama T, Ashiie T, Kikuri K, et al. *mitochondrial aspartate aminotransferase linked to IgG of the kappa-lambda type: report of a case*. *Clin Chim Acta* 1986; 160:297-305.
8. Litin SC, O' Brien JF, Pruett S, Foresman RW, Burritt MF, Bartholomew LG, Baldus WP *Macroenzyme as a cause of unexplained elevation of aspartate aminotransferase*. *Mayo Clin Proc* 1987; 62:681-687.
9. Foust RT, O'brien JF, Schiff ER. *Isolated aspartate aminotransferase elevation due to macroenzyme formation with liver biopsy correction*. *The American Journal of Gastroenterology Vol 85 No 1* 1990; 88-90.
10. Stasia MJ, Surla A, Renversez JC, Pene F, Femelez AM, Morel F. *Aspartate aminotransferase macroenzyme complex in serum identified and characterized*. *Clin Chem* 1994; 40:7 1340-1343.
11. Rej R, Vanderlinde RE, Fasce CF. *An L-aspartate: 2-oxoglutarate aminotransferase reference material from human erythrocyte: Preparation and characterization* *Clin Chem* 18/4 374-384 1978.
12. Sakakibara S, Shiomi K, Kobayashi S, Ikeda T, Inai S, Kagamiyama H. *A convenient and sensitive method for the determination of serum aspartate aminotransferase isoenzymes after electrophoresis*. *Clin Chim Acta* 1983; 133:119-123.

MACROASPARTASEMIA AS A CAUSE OF ISOLATED ELEVATION OF
ASPARTATE AMINOTRANSFERASE

13. Rej R. *Aspartate aminotransferase activity and isoenzyme proportions in human liver tissues. Clin Chem* 1978; 24/11 1971-1979.
14. Naples B, Vassault A, Charpin S, Lacour B, Berthelot P. *Serum mitochondrial aspartate aminotransferase as a marker of chronic alcoholism: Diagnostic value and interpretation in a liver unit. Hepatology* Vol. 6 No. 1986; 4:608-614.
15. Nagamine M, Okochi K. *Complexes of immunoglobulins A and G with aspartate aminotransferase isoenzymes in serum. Clin Chem* 1983; 29:379-381.
16. Rej R. *Measurement of aminotransferase: Part 1. Aspartate aminotransferase. CRC Crit Clin Lab Sci* 1984; 21:99-186.
17. Feldig P, Hytloft PH, Horder M. *The stability of blood, plasma and serum constituents during simulated transport. Scand J Clin Lab Invest* 1981; 41:35-40.
18. Niblock AE, Leung FY, Henderson AR. *Serum aspartate aminotransferase storage and the effect of pyridoxal phosphate* 1986; 108:461-465.
19. Fridhandler L, Berk JE. *Macroamylasemia (Review) Adv Clin Chem* 1978; 20:267-285.
20. Adachi K, Suzuki K, Ohno Y, Sato B. *Impaired amylase activities caused by binding of abnormal immunoglobulin A in patients with macroamylasemia. Clin Chim Acta* 1986; 154:103-113.
21. Wilkinson JH. *Clinical significance of enzyme activity measurements (Review). Clin Chem* 1970; 16:882-890.
22. Peltenburg HG, Hermans WT, Willems GM, Flendrig JG, Schmidt E. *Estimation of the fractional catabolic rate constants for the elimination of cytozolic liver enzymes from plasma. Hepatology* 1989; 10:833-839.
23. Schifferli JA, Roth P, Steiger G, Paccaud JP, Schimit M. *Macro-prostatic acid phosphatase in a patient's serum. Clin Chem* 1988; 34:2172-2174.
24. Stein W, Bohner J, Bahlinger M. *Macro-lipase - a new member of the family of immunoglobulin-linked enzymes. J Clin Chem Clin Biochem* 1987; 25:837-843.
25. Kohno H, Sudo K, Kanno T. *Intestinal alkaline phosphatase linked to immunoglobulin G of the kappa type. Clin Chim Acta* 1983; 135:41-48.
26. Klonoff DC. *Macroamylasemia and other immunoglobulin-complexed enzyme disorders. West J Med* 1980; 133:392-407.
27. Kajita Y, Majima T, Yoshimura M, et al. *Demonstration of antibody for glutamic pyruvic transaminase (GPT) in chronic hepatic disorder. Clin Chim Acta* 1978; 89:485-492.
28. Bohner J, Stein W, Kuhmann E, Eggstein M. *Serum creatine kinase BB linked to immunoglobulin G. Clin Chim Acta* 1979; 97:83-88.
29. Burlina A, Secchiero S, Bertorelle R, Plebani M, Zaninotto M. *Immunoglobulin A (lambda chains) conjugated with lactate dehydrogenase in serum. Clin Chem* 1987; 33:1085-1086.
30. Alan T. Remaley, Peter Wilding. *Macroenzymes: Biochemical characterization, clinical significance, and laboratory detection. Clin Chem* 1989; 35/12 2261-70.
31. Philip J. Galasso, Scott C. Litin, John F. O' Brien. *The macroenzyme: A clinical review. Mayo Clin Proc* 1993; 68:349-354.